

1 **Diaphragm Plasticity in Aging and Disease:**
2 **Therapies for Muscle Weakness go from Strength to Strength**

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25 **Introduction**

26 The diaphragm is the main inspiratory muscle and is required to be highly active
27 throughout the lifespan. The diaphragm muscle must be able to produce and sustain various
28 behaviors that range from ventilatory to non-ventilatory such as those required for airway
29 maintenance and clearance. Throughout the lifespan various circumstances and conditions may
30 affect the ability of the diaphragm muscle to generate force and in turn the diaphragm muscle
31 may undergo significant weakness and dysfunction. For example, hypoxic stress, critical illness,
32 cancer cachexia, chronic obstructive pulmonary disorder (COPD), and age-related sarcopenia
33 all represent conditions in which significant diaphragm muscle dysfunction exists.

34 This perspective review article presents several interesting topics involving diaphragm
35 plasticity in aging and disease that were presented at the International Union of Physiological
36 Sciences (IUPS) Conference in 2017. During the lecture respiratory physiologists gave
37 overviews of the function of the diaphragm muscle and the effects of various comorbidities on
38 the respiratory system. Furthermore each speaker presented current therapeutic options or
39 targets that have shown promise in mitigating the deleterious effects imposed on the diaphragm
40 muscle. Each section below summarizes individual presentations given during this lecture at
41 IUPS including a brief concluding summary for each section. Furthermore, a general concluding
42 paragraph is also given at the end of the review along with the future perspectives. The goal of
43 the lectures and this perspective review is to maximize the broad and collective research impact
44 on diaphragm muscle dysfunction, in the search for transformative treatment approaches to
45 improve the diaphragm muscle health throughout the lifespan (Fig. 1).

46

47 **Hypoxia-induced diaphragm dysfunction: Strengthening the case for antioxidant**
48 **intervention in respiratory patients**

49 **The diaphragm muscle: Plasticity in the pump of life**

50 The diaphragm is the primary muscle of breathing, pivotal in the breath-by-breath control
51 of respiratory homeostasis. Akin to other striated muscles, the diaphragm retains a remarkable
52 capacity for plasticity in health and disease. Intrinsic malleability in the structure-function
53 relationship of the diaphragm may confer dynamic flexibility supporting respiratory performance
54 in the course of physiological stressors such as development and exercise. Conversely,
55 respiratory muscle remodeling in response to pathophysiological stressors can have deleterious
56 consequences for respiratory mechanics, potentially perpetuating respiratory morbidity, for
57 example in disease. Arguably, diaphragm atrophy and weakness in the critical care setting is
58 the exemplar of egregious diaphragm muscle plasticity, with expedient and profound
59 consequences for physiological performance (49). It is also very well established that diaphragm
60 remodeling and dysfunction presents in several chronic respiratory diseases, perhaps most
61 notably in COPD (7). Moreover, sarcopenia and weakness are evident in the aged diaphragm
62 (38). Whatever the driver of dysfunction, impaired diaphragm performance is associated with
63 poor prognosis for patients.

64 Oxygen homeostasis is critical to diaphragm function. Yet, despite the clinical relevance,
65 the effects of hypoxia on the intrinsic properties of the diaphragm are under-explored and
66 consequentially relatively poorly understood. Oxygen deficit presents in many guises in
67 physiological and pathophysiological scenarios (e.g. exercise, altitude, surgery, disease) with
68 often considerably diverse paradigms of exposure. It is therefore unsurprising that hypoxia-
69 dependent plasticity in respiratory muscles can span a spectrum of change from phenotypic
70 responses that appear to confer at least compensatory (e.g., force preservation in the face of a
71 stressor) and perhaps improved measures of performance, to overt changes that are clearly
72 deleterious for muscle, and hence system performance (59, 76). Pattern, intensity and duration
73 of hypoxic exposure are key determinants of functional outcomes. Muscle-specific effects
74 related to intrinsic differences in structure, function, and metabolic signatures are also
75 recognized. Emerging evidence reveals that age and sex are also influential factors shaping

76 respiratory muscle responses to hypoxia, with evidence of muscle-specific early-life
77 susceptibility to hypoxia, and female advantage in adult animals due to protective effects of
78 estrogen. Phenotypic change in muscle in response to perturbations in oxygen availability, often
79 referred to as adaptive and maladaptive plasticity, reveals that hypoxia is much more than a
80 mere symptom of respiratory morbidity. Rather, hypoxic stress has a capacity to exert major
81 influence on respiratory muscle form and function with resultant consequences for respiratory
82 system performance. This short article provides an overview of studies in rodent models of
83 chronic hypoxia, which have provided insight to characteristics and mechanisms of hypoxia-
84 induced respiratory muscle plasticity. The findings may have relevance to human diaphragm
85 function at high altitude and in hypoxic respiratory disease.

86 **The hypoxic diaphragm: Adaptive plasticity**

87 There is evidence of apparent improved functional performance, termed adaptive or
88 compensatory plasticity in response to hypoxia in diaphragm muscle, responses which serve to
89 protect against oxygen deficit, or limit the deleterious consequences of hypoxic stress on
90 muscle performance (hypoxic tolerance). There appears to be relative resilience in rodent
91 diaphragm, compared with other muscles, in response to hypoxia, whether presented as a
92 chronic sustained exposure mimicking altitude and pulmonary disease, or as a chronic
93 intermittent stimulus modeling sleep-disordered breathing. Rat and mouse diaphragm fatigue
94 tolerance is preserved or increased in response to 4-6 weeks of sustained hypoxia, whereas
95 increased fatigue is revealed in other striated muscles (31, 32, 72). Chronic sustained hypoxia
96 increases diaphragm Na^+/K^+ ATPase pump content (72) and mitochondrial aerobic efficiency
97 (31), and results in myofiber atrophy with increased capillary length to fiber perimeter ratio (32),
98 responses peculiar it seems to diaphragm compared with other striated respiratory and non-
99 respiratory muscles (32, 72). Exposure to chronic sustained hypoxia also increases rat
100 diaphragm muscle tolerance of subsequent acute severe hypoxic stress, insofar as force-
101 generating capacity is better preserved during severe hypoxia in chronic hypoxic diaphragm

102 compared with control (58). It is important to note however, that fatigue is commonly assessed
103 as the time-dependent run down in force-generating capacity relative to pre-fatigue force.
104 However, basal force is often reduced following chronic hypoxic exposure, complicating the
105 interpretation of improved fatigue tolerance in the wider context of muscle performance. Early
106 life exposure to sustained hypoxic stress increases rat pharyngeal dilator muscle fatigue,
107 whereas diaphragm fatigue tolerance is preserved (14). A similar outcome is noted in early life
108 exposure to chronic intermittent hypoxia, with evidence of persistent pharyngeal dilator muscle
109 weakness, but maintenance of diaphragm force-generating capacity (70, 71). The few studies
110 that have explored this issue paint a portrait of an essential pump with properties that protect
111 from the insidious effects of hypoxia.

112 **The hypoxic diaphragm: Maladaptive plasticity**

113 Whereas adaptive plasticity is portrayed in rodent diaphragm muscle in response to
114 chronic hypoxia, contemporaneous maladaptive plasticity has also been revealed (59, 76).
115 Chronic hypoxia causes rat and mouse diaphragm weakness (61, 72), consistent with evidence
116 of diaphragm myofiber atrophy (31, 72). It is also established that chronic sustained hypoxia
117 decreases type IIa fiber specific force (20), revealing fiber dysfunction in addition to fiber
118 atrophy. The significance of these observations is that diaphragm weakness is recognized as
119 having prognostic value in the management of respiratory patients. Chronic sustained hypoxia
120 was shown to cause progressive and extensive protein oxidation, and metabolic remodeling
121 culminating in a p38 MAP kinase/FOXO-3a/20S proteasome-dependent atrophic response
122 causing diaphragm weakness in mice (61). Curiously, chronic intermittent hypoxia causes rat
123 diaphragm weakness and fatigue without overt oxidative stress (93), notwithstanding that other
124 studies have revealed oxidant stress following chronic intermittent hypoxia in muscle and other
125 tissues (59, 76). Deleterious effects of chronic sustained hypoxia and chronic intermittent
126 hypoxia extend to pharyngeal dilator muscles (62, 97), which could serve to further compromise
127 the integrative control of breathing. The experimental evidence points to differential outcomes

128 that depend upon pattern, duration and intensity of hypoxia, as well as age and sex (59, 60, 76,
129 77). The time-dependent elaboration of diaphragm dysfunction in response to sustained hypoxia
130 over 1-6 weeks of exposure reveals a temporal oxidative stress (61), due to progressive pro-
131 oxidant production and a loss of antioxidant capacity. Therefore the threshold or tipping point in
132 the transition from adaptation to maladaptation appears to be related to the oxidant:antioxidant
133 balance, with muscle dysfunction arising during net oxidation. Oxidation can increase or
134 decrease calcium sensitivity of the contractile apparatus, which might explain the capacity for
135 divergent outcomes in skeletal muscle performance in response to hypoxia-dependent oxidative
136 stress. S-glutathionylation of the fast troponin I isoform increases rat and human fast fibre
137 sensitivity to calcium (74), enhancing muscle performance. Conversely, excess H₂O₂ and
138 downstream hydroxyl radicals depress fast fibre calcium sensitivity, with endogenous
139 glutathione levels serving as an important gatekeeper of redox status and muscle performance
140 (75). Future studies should determine calcium sensitivity of the myofilaments in hypoxic
141 diaphragm, and establish the extent of oxidation in myofibrillar proteins.

142 It appears that physiological trade-offs are at play in respect of respiratory muscle
143 responses to hypoxic stress. Diaphragm weakness is a hallmark of hypoxic exposure perhaps
144 at the expense of cellular strategies providing homeostatic defense and improved tolerance of
145 oxygen lack. In the light of all considerations, the data suggest that hypoxia can perpetuate
146 respiratory morbidity through deleterious effects on diaphragm muscle performance.

147 **Antioxidant supplementation prevents hypoxia-induced diaphragm dysfunction**

148 Dietary supplementation with antioxidants ameliorates or completely prevents respiratory
149 muscle dysfunction following chronic intermittent hypoxia (79). N-acetyl cysteine
150 supplementation is the most effective intervention in preventing rat diaphragm weakness and
151 fatigue elaborated by chronic intermittent hypoxia (93). N-acetyl cysteine is also effective in
152 preventing chronic sustained hypoxia-induced airway dilator (62) and diaphragm (61)
153 dysfunction in mice. Antioxidant supplementation prevented respiratory muscle protein

154 carbonylation following 6 weeks of sustained hypoxia (61, 62). Moreover, mouse diaphragm
155 weakness following just 8 hours of exposure to sustained hypoxia (79) is reversed by N-acetyl
156 cysteine pre-treatment (78). A comparison of the effects of antioxidants on respiratory muscle in
157 animal models of hypoxic stress is provided in a recent review article (76). The findings of these
158 studies provide strong rationale for a comprehensive assessment of N-acetyl cysteine
159 supplementation intervention in human trials.

160 **Conclusions**

161 The multi-faceted stimulus of oxygen deficit can drive adaptive and maladaptive
162 outcomes for respiratory muscle performance. Hypoxia is a feature of high altitude and it
163 commonly presents as a consequence of respiratory morbidity, with a capacity whatever the
164 cause to contribute further to impaired respiratory performance and perhaps the spiral of
165 disability characteristic of chronic respiratory diseases. Altered redox signaling is pivotal in
166 driving diaphragm plasticity in response to hypoxic stress. The experimental evidence in animal
167 models clearly demonstrates the efficacy of N-acetyl cysteine supplementation in preventing
168 hypoxia-induced diaphragm dysfunction. On this basis, there is adequate justification to explore
169 the potential of adjunctive antioxidant therapies for respiratory muscle dysfunction in human
170 respiratory disease.

171

172 **Diaphragm fiber weakness in the critically ill**

173 **Diaphragm weakness in critically ill patients**

174 Patients with critical illness experience substantial skeletal muscle weakness and
175 physical disability. This leads to functional impairment of survivors of the Intensive Care Unit
176 (ICU), an impairment that can last for years (11, 47, 91). In particular, weakness of the
177 diaphragm – the main muscle of inspiration – is of major concern in critically ill patients: it
178 prolongs ventilator dependency, increases morbidity and duration of hospital stay, and it is
179 associated with long term functional limitations after hospital discharge (12, 19, 27, 35).

180 Diaphragm weakness in mechanically ventilated critically ill patients has been established with
181 non-invasive measurements; ultrasound revealed reduced motion and thinning of the diaphragm
182 (22, 36, 44, 54), and by magnetic stimulation of the phrenic nerves a reduced capacity to
183 generate pressure was observed (21, 35, 46, 52). The cellular changes that underlie diaphragm
184 weakness in critically ill patients are unclear. Changes in phrenic nerve function, in
185 neuromuscular transmission, or in the contractility of individual muscle fibers all may explain the
186 reduction in pressure generation by the diaphragm.

187 A plethora of data suggests that changes in the contractility of individual diaphragm
188 muscle fibers play a major role. Critical illness-associated phenomena, such as mechanical
189 ventilation-induced diaphragm inactivity (55, 90, 92), malnutrition (57), and inflammation (86)
190 are associated with contractile weakness of diaphragm fibers and activation of proteolytic
191 pathways within diaphragm fibers in animal models. Whether these findings translate to humans
192 is unknown, although several studies (51, 56), but not all (50), in brain dead organ donors who
193 received mechanical ventilation prior to organ harvest revealed atrophy and activation of the
194 ubiquitin-proteasome pathway in diaphragm muscle fibers. However, brain dead organ donors
195 do not exhibit the clinical features of critically ill patients; complete absence of neural activation
196 of the diaphragm, metabolic stress and brain ischemia differentiates them. Consequently, it is
197 unknown whether these findings translate to critically ill patients.

198 **Studies on individual diaphragm fibers of critically ill patients**

199 To fill this void, recent studies aimed to study the contractile function of diaphragm fibers
200 of critically ill patients. We obtained diaphragm biopsies of critically ill patients who received
201 mechanical ventilation prior to surgery, and compared these biopsies with those obtained from
202 patients undergoing resection of an early lung malignancy (controls). The size and the
203 contractile strength of individual muscle fibers were determined. Both slow-twitch and fast-twitch
204 diaphragm muscle fibers of critically ill patients had a ~25% smaller cross sectional area, and a
205 >50% lower absolute contractile force (49), (note that the contractile properties of individual

206 diaphragm fibers depend on fiber type (33, 34), and therefore data from slow- and fast-twitch
207 fibers are reported separately). Additionally, we evaluated critical components of the ubiquitin-
208 proteasome pathway, and performed proof-of-concept studies in MuRF-1 knockout mice to
209 evaluate the role of this pathway in the development of contractile weakness during mechanical
210 ventilation (49). Markers of the ubiquitin-proteasome pathway, in particular MuRF-1, a muscle –
211 specific ubiquitin ligase, were significantly upregulated in the diaphragm of critically ill patients.
212 MuRF-1 deficient mice were protected against the development of diaphragm contractile
213 weakness during mechanical ventilation, indicating that MuRF-1 upregulation in the diaphragm
214 of critically ill patients might play an important role in the development of fiber atrophy and
215 contractile weakness.

216 The mechanisms that trigger proteolysis in diaphragm muscle fibers of critically ill
217 patients are incompletely understood. Studies on animal models and on brain dead organ
218 donors suggest that these mechanisms include oxidative stress, induced by mitochondrial
219 alterations during mechanical ventilation (56, 69, 80). For instance, the influx of energetic
220 substrates, such as lipid and glucose, might exceed the demand of diaphragm muscle fibers
221 during a sudden drop in substrate utilization rate during mechanical unloading of the diaphragm
222 in brain dead organ donors and mechanically ventilated rodents (80, 81). This state of metabolic
223 oversupply then promotes mitochondrial fission, mitochondrial dysfunction and increase
224 oxidative stress (53). Surprisingly, recent studies from our group on thirty-six mechanically
225 ventilated critically ill patients, whom displayed significant atrophy and contractile weakness of
226 diaphragm muscle fibers, revealed absence of mitochondrial dysfunction, oxidative stress, and
227 metabolic oversupply in diaphragm fibers (98). These findings suggest that mitochondria and
228 redox status should not be the primary target of therapy aimed at preserving diaphragm function
229 in critically ill patients.

230 **Targeting the diaphragmatic sarcomere**

231 Recent studies tested the ability of troponin activators to improve diaphragm fiber
232 contractility in critically ill patients. These compounds target the sarcomere, the smallest
233 contractile unit in muscle, and in particular those in fast-twitch muscle fibers. These fast-twitch
234 diaphragm fibers of critically ill patients were of particular interest as they not only have reduced
235 maximal force, but they also require more calcium to generate submaximal force (48). To test
236 whether this reduced calcium-sensitivity of force can be restored, diaphragm fibers of critically ill
237 patients were exposed to the fast skeletal troponin activator CK-2066260. Fast skeletal troponin
238 activators increase the affinity of the troponin complex to Ca^{2+} (87). Compared to vehicle, 5 μ M
239 of CK-2066260 significantly increased the calcium sensitivity of diaphragm fibers, both in
240 controls and in critically ill patients. Importantly, at physiological calcium concentrations, CK-
241 2066260 restored the contractile force of fast-twitch diaphragm fibers of critically ill patients back
242 to levels observed in untreated fibers from controls (48).

243 **Conclusions**

244 To date, no drug is approved to improve respiratory muscle function in mechanically
245 ventilated critically ill patients. Administration of troponin activators might prove an elegant
246 strategy to restore diaphragm fiber strength. These compounds improve contractility at calcium
247 concentrations which reflect activation during daily live activities (87). And since ~50% of fibers
248 and total fiber area in the human diaphragm consists of fast-twitch fibers, fast skeletal troponin
249 activators might significantly improve in vivo diaphragm strength. Similarly, studies with
250 levosimendan, a commercially available calcium sensitizer that targets sarcomeric troponin in
251 slow-twitch muscle fibers, showed improved neuromechanical efficiency and contractile function
252 of the diaphragm in healthy controls (25). These findings underscore the therapeutic potential of
253 troponin activators.

254

255 **Age-related sarcopenia in the diaphragm muscle**

256 **Aging and sarcopenia**

257 The aging population is increasing steadily both in the USA and worldwide. In the USA
258 alone, there are currently 48 million people over the age of 65, and this is expected to increase
259 to about 88 million by 2050 (45). Aging alone is related to increased incidence of chronic
260 diseases, and a single intrinsic risk factor underlying many disease states. For example,
261 cardiovascular disease and osteoporosis have a significantly increased prevalence with age.
262 Sarcopenia is the age-related disorder of skeletal muscle, and is associated with the loss of
263 muscle mass and function (30). To date, sarcopenia has been well defined and investigated in
264 limb muscles but less so in the diaphragm muscle (13). In a series of studies we established the
265 effects of diaphragm muscle sarcopenia (26, 38, 39, 41), investigated mechanistic drivers (37,
266 42), and evaluated possible therapeutic approaches to target age-related changes to the
267 diaphragm muscle (43).

268 The two main components of sarcopenia were investigated in the diaphragm muscle of
269 mice at ages of 100%, 90%, and ~75% survival (38, 39, 41). The first component investigated
270 was the force generating capacity of the diaphragm muscle. The old mice at 75% survival had
271 ~34% less force generating capacity (i.e., maximal specific force) than young mice (100%
272 survival) and those in early old age (90% survival). Significant force loss occurred in mice
273 between 90 and 75% survival, a window of only six months (18 – 24 months of age). Loss of
274 maximal diaphragm muscle force is expected to impact the ability to perform non-ventilatory
275 behaviors related to airway clearance. The second component of sarcopenia investigated was
276 the type specific cross-sectional area of diaphragm muscle fibers. There is a loss of muscle fiber
277 size of the type IIx and/or IIb muscle fibers, but not in type I and IIa diaphragm muscle fibers
278 between 100% and 75% survival in mice (38, 41). Collectively this extent of sarcopenia is
279 expected to significantly impair the ability of the diaphragm muscle to accomplish a broad range
280 of behaviors in old age, and limit the ability to maintain airways clear. The overarching goal of
281 this on-going work has been to understand if sarcopenia is related to trophic changes

282 throughout the motor unit and to evaluate mitigating approaches for age-related diaphragm
283 neuromuscular transmission failure and sarcopenia.

284 **Diaphragm motor unit recruitment**

285 Motor units are composed of an individual motor neuron that innervates a group of
286 muscle fibers through a neuromuscular junction on each fiber, and the motor unit is known as
287 the basic unit of neuromotor control (63). Individual motor units are classified as fast-twitch (type
288 F) and slow-twitch (type S) based on specific mechanical and fatigue properties of the muscle
289 fibers. The type F motor units are further categorized into fast-twitch fatigue resistant (type FR),
290 fast-twitch fatigue intermediate (type FInt), and fast-twitch fatigable (type FF) based on specific
291 metabolic capacities of the muscle fibers. The type FInt and FF motor units comprise type IIX
292 and IIb muscle fibers, primarily expressing myosin heavy chain MyHC_{2X} and MyHC_{2B},
293 respectively, which are the fibers that display the preferential age-related loss in cross-sectional
294 area.

295 The diaphragm muscle must be able to accomplish ventilatory behaviors such as normal
296 room air breathing (eupnea) and during stimulation from increased chemical drive (hypoxia-
297 hypercapnia or exercise). Additionally, the diaphragm muscle must also perform non-ventilatory
298 behaviors that require greater force and are related to maintenance of airway patency and
299 airway clearance (e.g., sneezing, gagging and coughing). Using a well-established model of
300 diaphragm muscle neuromotor control across a range of diaphragm behaviors (94, 96) it is
301 known that ventilatory behaviors require only the type S and FR motor units, while non-
302 ventilatory behaviors also require type FInt and FF (66, 67, 95). Knowing that diaphragm muscle
303 sarcopenia selectively atrophies the more forceful type IIX and/or IIb fibers, we investigated if
304 there was an age-related impairment in maximal behaviors using transdiaphragmatic pressure
305 (Pdi) measurements, a surrogate for diaphragm muscle force, in mice at ages of 100% and
306 ~75% survival (39, 40). Consistent with the large reserve capacity for force generation by the
307 diaphragm muscle, aging did not affect the ability to generate forces necessary for ventilatory

308 behaviors, however there was significant age-related impairment of more forceful non-
309 ventilatory behaviors.

310 **Trophic influences**

311 The exact mechanisms responsible for diaphragm muscle sarcopenia remain unclear,
312 but it is possible that they may be related to changes throughout the neuromuscular system.
313 The first step in understanding age-related trophic changes was to evaluate effects at the
314 neuromuscular junction. In a series of studies, we investigated neuromuscular transmission
315 using a global measure of force generation by the diaphragm muscle in response to nerve and
316 direct muscle activation during aging. In dissimilarity to aging effects on maximal force
317 generation, alterations in diaphragm neuromuscular transmission were evident in mice by early
318 old age with no further change into old age (90 vs 75% survival, respectively) (37). These
319 findings indicate that the changes in neuromuscular transmission precede the loss of diaphragm
320 muscle force or significant muscle fiber atrophy.

321 There are various possible trophic influences at the neuromuscular junction and it is
322 understood that neurotrophins can acutely enhance neuromuscular transmission. It is unclear if
323 age-related alterations in neurotrophins could help to explain the detrimental effects of
324 sarcopenia on neuromuscular transmission. The neurotrophin brain-derived neurotrophic factor
325 (BDNF) acting through its high affinity tropomyosin-related kinase receptor subtype B (TrkB)
326 receptor is known to play an important role in the development and maintenance of adult
327 neuromuscular junctions and has an important role in neuromuscular transmission (65, 68).
328 Furthermore, in young diaphragm muscles it is known that enhancing BDNF signaling can
329 improve neuromuscular transmission, while inhibition of TrkB kinase activity can impair
330 neuromuscular transmission (68).

331 To investigate possible age-related alterations in trophic interactions throughout the
332 motor unit we investigated the effects of BDNF/TrkB signaling in the aging diaphragm muscle
333 and neuromuscular junction. In early old age (~90% survival), increased BDNF improved

334 neuromuscular transmission, while inhibition of TrkB kinase activity had no effect. However, in
335 old age (~75% survival) neither increased BDNF nor the inhibition of TrkB kinase activity had
336 any effect on diaphragm neuromuscular transmission. This suggested that there may be
337 reductions in endogenous BDNF in the aging diaphragm muscle which likely preceded
338 reductions in TrkB kinase activity (37). Second, examining the early old age time point, given
339 that neuromuscular transmission was impaired without evidence of sarcopenia, a detailed
340 morphological investigation of the neuromuscular junction was conducted. Using a knockin
341 *TrkB*^{F616A} mouse model allowing for reversible inhibition that is sensitive to the phosphoprotein
342 phosphatase-1 derivative (1NMPP1) (18) BDNF/TrkB signaling was disrupted for one week. The
343 inhibition of TrkB kinase activity in early old age increased the proportion of denervated
344 neuromuscular junctions in the diaphragm muscle (42). Collectively, the early old age time point
345 indicated loss of neuromuscular transmission and a period of susceptibility during early old age
346 in which BDNF/TrkB signaling at diaphragm neuromuscular junctions supports the maintenance
347 of neuromuscular junctions structure and muscle innervation, prior to the onset frank
348 sarcopenia.

349 In efforts to mitigate both diaphragm muscle sarcopenia and neuromuscular
350 transmission failure, BDNF/TrkB signaling was targeted therapeutically starting at early old age.
351 Previously, in young adult mice the highly selective BDNF analog and TrkB agonist, 7,8-
352 dihydroxyflavone (7,8-DHF) was shown to acutely improve diaphragm neuromuscular
353 transmission. As such, chronic treatment was investigated starting at early old age for a six
354 month period. However, contrary to the anticipated effects, chronic treatment with 7,8-DHF was
355 not able to mitigate diaphragm muscle sarcopenia or impairments in neuromuscular
356 transmission (43). While the therapeutic approach with 7,8-DHF was not effective in mitigating
357 age-related dysfunction it is possible that targeting BDNF/TrkB signaling may still be an effective
358 treatment approach if started earlier.

359 **Conclusions**

360 Our work has sought to understand neuromuscular dysfunction in the aging diaphragm
361 muscle by understanding the impact of age-related impairments in BDNF/TrkB signaling. The
362 most salient findings of this on-going work have been the evidence for a time course of age-
363 related changes in neuromuscular activity and trophic signaling. The reduction in BDNF
364 availability likely precedes a loss of TrkB receptor expression and occurs when there are
365 reversible changes in neuromuscular transmission but not yet evidence of sarcopenia. This
366 work supports ongoing investigations of the mechanisms underlying disrupted trophic signaling
367 at the neuromuscular junction and highlights the importance of understanding the role of motor
368 neuron and neuromuscular junction dysfunction in the pathogenesis of sarcopenia.

369

370 **Diaphragm dysfunction in cancer cachexia: mechanisms and therapies**

371 **Skeletal muscle dysfunction in chronic respiratory diseases**

372 In patients with chronic respiratory conditions, dysfunction of the ventilatory and limb
373 muscles is frequently observed as a relevant systemic manifestation of these diseases. Other
374 conditions such as chronic heart failure, cancer cachexia, and sepsis may also induce both
375 respiratory and peripheral muscle dysfunction and mass loss in patients as well as in animal
376 models (2, 3, 6, 64, 73, 99). Skeletal muscle dysfunction is of multifactorial etiology . Several
377 factors intrinsic to both the host (whether patient or animal) and the condition itself and
378 biological mediators interact together to alter the function of the muscles characterized by a
379 decline in either the strength or endurance properties. Interestingly, the mechanical factors,
380 mainly due to the increased inspiratory loads of patients with chronic respiratory diseases,
381 contribute to a great extent to the ventilatory muscle dysfunction described in these patients,
382 while those factors are irrelevant in the etiology of the dysfunction of locomotor muscles.

383 The most relevant factors and biological mediators that contribute to respiratory and limb
384 muscle dysfunction in patients and animal models have been extensively reviewed in several
385 review articles (2, 3, 6, 64, 73, 99). Briefly, factors such as hypoxia, hypercapnia, acidosis,

386 cigarette smoking, metabolic disorders, nutritional abnormalities, aging, genetic predisposition,
387 drugs, comorbidities, systemic inflammation, and inactivity have been shown to contribute to
388 muscle mass loss and impaired function in animal models and patients (2, 3, 6, 64, 73, 99). In
389 the respiratory muscles, mainly the diaphragm, the contribution of the alterations in the thorax
390 geometry and the increased inspiratory loads to which chronic respiratory patients are
391 continually exposed, are major contributors to the reported ventilatory muscle dysfunction,
392 especially in the initial stages (2, 3, 6, 64, 73, 99). Furthermore, biological mediators such as
393 oxidative stress, increased protein breakdown and muscle protein degradation, poor anabolism,
394 mitochondrial alterations, and epigenetic events are counted among the most relevant
395 mechanisms shown to be involved in the skeletal muscle dysfunction of the patients and animal
396 models (2, 3, 6, 28, 29, 64, 73, 82-85, 99). Moreover, in the early stages of respiratory muscle
397 dysfunction, several biological adaptations take place in the diaphragm muscle as a result of the
398 increased inspiratory loads intrinsic to chronic respiratory disease (3, 6). A rise in capillary
399 contacts, shorter sarcomere length, increased oxidative capacity, slow-twitch fiber proportions,
400 in myoglobin and in mitochondrial content are the most relevant adaptive mechanisms
401 described in the diaphragm of patients with chronic respiratory conditions (3, 6). The adaptive
402 mechanisms counterbalance the deleterious effects inherent to the host and disease up to a
403 certain point. When the harmful conditions (e.g., hypoxia and increased metabolic and oxygen
404 demands) outweigh the adaptive scenario, respiratory muscle dysfunction occurs in the patients
405 with chronic respiratory conditions (3, 6).

406 **Respiratory muscle dysfunction: Biological mechanisms and therapeutic targets in** 407 **several conditions**

408 **Sepsis.** A decline in diaphragm force generation has been consistently observed in
409 endotoxemic mice and rats in many different studies. Our group demonstrated that heme
410 oxygenase activity is involved in muscle performance in normal and endotoxemic rats as its
411 selective inhibition with chromium-mesoporphyrin IX induced a further decline in muscle force

412 generation of diaphragm muscle strips (4). The antioxidant N-acetyl cysteine was shown to
413 induce an improvement in respiratory muscle function (*in vivo* measurements) in septic rats
414 together with a rise in superoxide dismutase activity and a decline in diaphragm oxidative stress
415 (10).

416 **Cigarette smoking models.** Chronic exposure to cigarette smoking induced a
417 significant rise in protein oxidation levels, especially of creatine kinase, in the diaphragm of
418 guinea pigs (8) and mice (5). Moreover, it also induced a significant decline in body weight gain
419 over time in both animal models, suggesting that the compounds of cigarette smoke may
420 influence body compartments and composition in these animal models (5, 8).

421 **Hypoxia.** Exposure to chronic intermittent hypoxia also induced a decline in body weight
422 gain in rats after two weeks (23). Levels of proinflammatory cytokines were greater in the
423 diaphragm and plasma of the animals exposed to chronic intermittent hypoxia for 15 days (23).
424 Moreover, levels of superoxide anion were also significantly increased in the mitochondria and
425 membrane compartments of the diaphragm muscle and plasma of the rats exposed to the 15-
426 day period of chronic intermittent hypoxia (23). Treatment of the animals with the antioxidant N-
427 acetyl cysteine induced a significant reduction in superoxide anion levels as measured by
428 chemiluminescence in both diaphragm muscle and systemic levels of the rats exposed to
429 hypoxia compared to non-exposed animals (23). Treatment with the anti-tumor necrosis factor
430 (TNF)-alpha antibody of the rats exposed to chronic intermittent hypoxia reduced levels of
431 superoxide anion in the diaphragm muscle compartments (membrane and mitochondria) and
432 plasma, while it only induced a significant decline in the systemic levels of proinflammatory
433 cytokines, but not in muscles (23).

434 **Inspiratory threshold loading models.** The diaphragm was also analyzed in rats
435 exposed to several degrees of inspiratory threshold loading in another investigation(23, 24). A
436 decline in body weight gain was observed in the rats exposed to the greatest loads (70% of
437 maximal inspiratory pressure, MIP) (24). Superoxide anion levels were greater in the

438 mitochondrial and membrane compartments of the diaphragm of the 70%-MIP rats and
439 treatment with either N-acetyl cysteine or anti-TNF-alpha antibody induced a significant decline
440 in those levels even below control levels (24).

441 **Chronic heart failure.** Experimental chronic heart failure (CHF) induced as a result of a
442 single dose of monocrotaline injection in rats, which experienced a significant reduction in body
443 weight gain and muscle mass loss (9). Treatment with either bortezomib (proteasome inhibitor)
444 or N-acetyl cysteine attenuated these deleterious effects on the animals after one month (9).
445 Levels of proteolysis (tyrosine release) were also increased in the diaphragm and limb muscles
446 of the CHF rats and treatment with bortezomib and N-acetyl cysteine significantly reduced those
447 levels (9).

448 **Diaphragm dysfunction in cancer-induced cachexia and potential therapies**

449 Several therapeutic strategies to treat muscle wasting in models of cancer-induced
450 cachexia have been tested in animals in a great variety of studies. In this regard, in this section
451 the effects of different therapeutic agents will be subsequently described in experimental models
452 of lung cancer-induced cachexia.

453 **Models of lung cancer carcinogenesis in mice.** Our group has published several
454 studies focused on the elucidation of the underlying biological mechanisms of muscle mass loss
455 in respiratory and limb muscles in animal models of cancer cachexia. Body weight gain and
456 muscle mass were significantly reduced in mice exposed to chronic lung carcinogenesis with
457 urethane for several time-points (88). Increased muscle injury and apoptosis along with a
458 decline in the content of specific structural and functional muscle proteins were seen in the
459 diaphragm of the cancer-induced cachectic mice compared to non-exposed control animals
460 (88).

461 **Models of lung cancer cell inoculation in wild type mice.** In another model of lung
462 cancer cell inoculation in mice, treatment with several agents such as specific inhibitors of
463 nuclear factor (NF)- κ B and mitogen-activated protein kinases (MAPK) pathways, but not

464 bortezomib, attenuated the loss in body weight gain and diaphragm and gastrocnemius muscle
465 mass and reduced limb strength in the animals (16). Levels of proteolysis (tyrosine release)
466 were attenuated in the diaphragm and limb muscles in response to treatment with either the
467 proteasome, MAPK, or NF-κB inhibitors after one month of study (16). Cachexia-induced NF-κB
468 signaling was also attenuated as a result of treatment of the animals with either bortezomib for
469 the corresponding NF-κB inhibitor (16). In the same animal model of lung cancer-induced
470 cachexia, a decline in the mitochondrial respiratory chain complexes I and IV and oxygen
471 consumption was observed in the diaphragm and gastrocnemius of the cachectic mice
472 compared to control non-cachectic animals (28). Importantly, treatment with either NF-κB or
473 MAPK inhibitors improved, especially in the diaphragm, mitochondrial respiratory chain activity
474 and oxygen consumption in the cachectic mice (28). The proteasome inhibitor bortezomib,
475 however, did not elicit any significant improvement in those parameters in the cancer cachectic
476 mice (28). Lately, our group has also demonstrated that the beta₂ agonist formoterol exerted its
477 beneficial through reduced oxidative stress (89) and atrophy signalling levels (1) in both
478 diaphragm and especially the gastrocnemius muscle in rats with cancer-induced cachexia.

479 **Model of lung cancer cell inoculation in genetically deficient mice.** The effects of
480 poly(ADP-ribose) polymerases (PARP) on the process of muscle mass loss and wasting in
481 cancer-induced cachexia has also been recently explored. In fact, PARP activity was increased
482 in the diaphragm and gastrocnemius of the cancer-induced cachexia mice compared to the non-
483 cachectic control animals (17). Interestingly, in cancer cachectic mice deficient for either Parp-1
484 or Parp-2 genes, the decrease in total body weight gain, muscle weights, and limb strength was
485 attenuated compared to cachectic wild type animals (15, 17). Moreover, the alterations
486 observed in several epigenetic mechanisms, namely reduced muscle-specific microRNA
487 expression and histone deacetylase levels, of the diaphragm and gastrocnemius muscles in the
488 *Parp1*^{-/-} and *Parp2*^{-/-} cachectic mice were also partially attenuated compared to those detected
489 in the wild type cachectic rodents (15). In the same model of PARP-1 and PARP-2 deficient

490 mice, the increased levels of oxidative stress, proteolysis (tyrosine release assay), ubiquitin-
491 proteasome pathway markers, and atrophy signaling pathways were also attenuated in the
492 cancer cachectic animals compared to the cachectic wild type mice (17). Additionally, the
493 reduction in myosin protein levels detected in the diaphragm of cachectic wild type animals was
494 also attenuated particularly in the *Parp2*^{-/-} cachectic mice (17).

495 **Conclusions**

496 In conclusion, several therapies have been demonstrated to be very efficient for the
497 treatment of muscle wasting in several models of cachexia including cancer. The antioxidant N-
498 acetyl cysteine and inhibitors of atrophy signaling pathways have shown an important
499 attenuation of the deleterious effects leading to muscle mass loss. Moreover, PARP activity
500 seems to play a relevant role in the process of muscle wasting, which could also be the basis for
501 the use of PARP inhibitors in models of cancer-induced cachexia that could eventually be
502 translated into the clinics.

503

504

505 **Conclusions and Future Directions**

506 The goal of this IUPS symposium and perspective review was to highlight our collective
507 research efforts to understand novel approaches for various forms of diaphragm muscle
508 dysfunction (Fig. 1). Collectively the results presented at IUPS indicate that while no specific
509 treatment option is currently approved to target global diaphragm muscle dysfunction, there are
510 several therapeutic strategies underway. Specifically, therapies have been demonstrated to be
511 very efficient for the treatment of various models of diaphragm muscle dysfunction.

512 First, the antioxidant N-acetyl cysteine and inhibitors of atrophy signaling pathways have
513 shown an important attenuation of the deleterious effects leading to muscle mass loss. Second,
514 PARP activity seems to play a relevant role in the process of muscle wasting, which could also
515 be the basis for the use of PARP inhibitors in models of cancer-induced cachexia that could

516 eventually be translated into the clinics. Third, targeting of calcium concentrations via troponin
517 activators may be transformative to critically ill ventilator-dependent patients. Forth, altered
518 redox signaling plays a pivotal in driving diaphragm plasticity in response to hypoxic stress. The
519 experimental evidence in animal models clearly demonstrates the efficacy of antioxidant
520 supplementation in preventing hypoxia-induced diaphragm dysfunction. Finally, in models of
521 aging currently understood disruptions in trophic signalling provide a possible target for novel
522 therapies although no specific therapies have been identified yet.

523 At present more research is needed in order to move these possible therapies to the
524 clinic. Furthermore, work to identify additional novel therapeutic targets may help mitigate the
525 deleterious effects of the loss in diaphragm function in chronic respiratory diseases, cancer as
526 well as in critical illness and aging.

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868 **Figure Legend**

869 **Figure 1.** Framework of the collective research impact on diaphragm muscle dysfunction
870 resented at the IPUS 2017. This symposium sought to understand possible treatment
871 approaches and targets to improve the diaphragm muscle health throughout the lifespan and
872 across chronic diseases and disorders. Throughout this perspective review we present various
873 biological factors, markers of dysfunction, and positive adaptations to potential treatments in
874 models of hypoxic stress, critical illness, cancer cachexia, chronic obstructive pulmonary
875 disorder (COPD), and age-related sarcopenia all of which present significant diaphragm muscle
876 dysfunction.

877 BDNF/TrkB: brain-derived neurotrophic factor / tropomyosin-related kinase receptor subtype B

878 PARP: poly(ADP-ribose) polymerases

879 TNF- α : tumor necrosis factor-alpha

880

881

882

Pathophysiologic Factors and Risks

Aging
Comorbidities
Environmental Factors & Cigarette Smoke
Genetics
Hypoxic Stress
Inactivity
Injury
Inspiratory Load
Malnutrition
Metabolic Stress
Oxidative Stress
Physical Disability
Sex
Systemic Inflammation

Markers of Dysfunction

Fatigue
Fiber Atrophy
Impaired Neuromuscular Transmission
Metabolic Remodeling
Mitochondrial Loss
Muscle Weakness
Protein Oxidation

Disorders Targeting the Diaphragm Muscle
Cancer Cachexia
Critical Illness
Chronic Obstructive Pulmonary Disease (COPD)
Hypoxic Stress
Sarcopenia
Cancer

Plasticity

Potential Treatments & Targets
Antioxidants
PARP Signaling
TNF- α Signaling
Trophic (BDNF/TrkB) Signaling
Troponin/Calcium Signaling
Ubiquitin-Proteasome Pathway
Beta₂ agonists (formoterol)

Documented Positive Adaptations

Hypoxic Tolerance
Improved Fatigue Tolerance
Improved Force Generating Capacity
Mitigation of Fiber Atrophy
Mitigation of Fiber Weakness