New	insights	into	the	molecular	pathophysiology	of	fragile	X	syndrome	and
therapeutic perspectives from the animal model.										

Arnau Busquets-Garcia§, Rafael Maldonado, Andrés Ozaita\*

Departament de Ciències Experimentals i de la Salut. Universitat Pompeu Fabra, 08003 Barcelona, Spain.

§Present address: INSERM U862 NeuroCentre Magendie, Bordeaux, France; Université de Bordeaux, Bordeaux, France.

# \*Corresponding author:

Andrés Ozaita, Laboratori de Neurofarmacologia, Facultat de Ciències de la Salut i de la Vida, Universitat Pompeu Fabra, Parc de Recerca Biomèdica de Barcelona, C/Doctor Aiguader 88, 08003 Barcelona, Spain. Phone: +34-93-3160823; Fax: +34-93-3160901; e-mail: andres.ozaita@upf.edu

### **Abstract**

Fragile X syndrome is the most common monogenetic form of intellectual disability and is a leading cause of autism. This syndrome is produced by the reduced transcription of the fragile X mental retardation (*FMRI*) gene, and it is characterized by a range of symptoms heterogeneously expressed in patients such as cognitive impairment, seizure susceptibility, altered pain sensitivity and anxiety. The recent advances in the understanding of the pathophysiological mechanisms involved have opened novel potential therapeutic approaches identified in preclinical rodent models as a necessary preliminary step for the subsequent evaluation in patients. Among those possible therapeutic approaches, the modulation of the metabotropic glutamate receptor signaling or the GABA receptor signaling have focused most of the attention. New findings in the animal models open other possible therapeutic approaches such as the mammalian target of rapamycin signaling pathway or the endocannabinoid system. This review summarizes the emerging data recently obtained in preclinical models of fragile X syndrome supporting these new therapeutic perspectives.

*Keywords:* autism, fragile X syndrome, mGluR5, mammalian target of rapamycin (mTOR), endocannabinoid system, CB1 cannabinoid receptor, intellectual disability, anxiety, epilepsy, nociception.

### Introduction

Fragile X syndrome (FXS) is the leading inherited neurological disorder causing intellectual disability and autism (Kooy et al., 2000). The causative mutation of almost all known cases of FXS is a trinucleotide CGG expansion in the promotor region of the fragile X mental retardation gene (*FMR1*) producing the loss of the fragile X mental retardation protein (FMRP) (Krueger and Bear, 2011; Penagarikano et al., 2007). FMRP is a RNA-binding protein that plays a major role in protein synthesis in neurons (Darnell et al., 2011).

The clinical symptoms of FXS are heterogeneous (Tranfaglia, 2011; Jacquemont et al., 2013). Although the most prominent neurological phenotype is intellectual disability (Fisch et al., 2002), patients with FXS often exhibit other neurodevelopmental problems, including attention deficit hyperactivity disorder and autistic-like behavior (Hagerman, 2006). Additional features of FXS include epilepsy, self-injurious behavior, sleep disorders and hypersensitivity to sensory stimulation (Chudley and Hagerman, 1987; Hagerman, 2006; Cornish et al., 2008).

Several cellular and molecular alterations have been demonstrated in FXS at the central nervous system level (Wang et al., 2012; Santoro et al., 2012). The most prominent is the alteration in the dendritic spine density and morphology (Hinton et al., 1991). This trait has also been observed in several autism spectrum disorders (Zoghbi, 2003; Spooren et al., 2012) and other neurological disorders (Penzes et al., 2011) leading to the idea that approaches that normalize this trait in FXS might also be worth tested in other brain disorders.

### Animal models of the disease with preclinical implications

The most studied animal model for FXS was obtained by interrupting the murine

*Fmr1* gene (Bakker, 1994) (Fmr1KO) that causes the loss of FMRP production. This animal model reproduces many aspects of the syndrome at the behavioral, cellular and molecular levels, allowing the evaluation of potential novel therapeutic approaches in the normalization of these pathological features.

#### Behavioral alterations observed in the animal model of FXS

The behavioral phenotype has been mainly characterized in the Fmr1KO mouse (Bakker, 1994). We will focus our attention to four of the behavioral traits better characterized: learning and memory impairment, seizure susceptibility, anxiety-like behavior and nociceptive desensitization, all sensitive to pharmacological or genetic intervention in the Fmr1KO.

Consistent with the cognitive impairment detected in FXS patients in attention-dependent tasks, the animal model shows clear deficits in the object recognition memory test (Ventura, 2004; Busquets-Garcia, 2013; Bhattacharya, 2012). Other tests demonstrate mild alterations such as those on spatial learning and reversal learning assessed in the Morris or plus-shaped water maze test (D'Hooge et al., 1997; Van Dam et al., 2000), or those on associative learning (Hayashi et al., 2007; Michalon et al., 2012; Paradee et al., 1999). These deficits have been also observed in the radial maze (Mineur et al., 2002). Interestingly, the deficits in spatial and associative learning seem to depend strongly on the strain where the Fmr1 mutation was studied (Paradee et al., 1999; Dobkin et al., 2000; Baker et al. 2010), an observation that reveals the critical role of the genetic background in certain behavioral outcomes.

(Musumeci et al., 2000). FXS patients may also respond to olfactory, tactile, visual, and auditory stimuli with hyper-reactivity and convulsions (Miller et al., 1999; Berry-

Kravis et al., 2010), and the prevalence of epilepsy in FXS is larger than in the normal population (Berry-Kravis et al., 2010). In this regard, the abnormally high synchrony and hyperexcitability in Fmr1KO mice cortical networks may explain the hypersensitivity and the predisposition to seizures, since normal circuit function requires precise and efficient excitatory and inhibitory neurotransmission balance (Gonçalves et al., 2013). Indeed, hippocampal neuronal circuits in the Fmr1KO model are epileptogenic, and cortical network activity is enhanced due to enhanced group I metabotropic receptor activity in these mice, both as a consequence of the enhanced group I metabotropic glutamate receptor signaling (Chuang et al., 2005; Havs et al., 2011), and the reduced interneuron activity (Paluszkiewicz et al., 2011). The appearance of audiogenic seizures is a robust and reproducible phenotype of interest to evaluate the potential impact of therapies in FXS (Michalon et al., 2012; Busquets-Garcia et al., 2013; Dolan et al., 2013; Pacey et al., 2009). Notably, hyperexcitable networks have been observed in other autistic syndromes (Markram and Markram, 2010), and in this respect approaches that reduce circuit hyperexcitability may become common therapeutic strategies to different pathologies (Wondolowski and Dickman, 2013; Eichler and Meier, 2008).

Anxiety is another common symptom displayed by FXS patients (Bailey et al., 2008). Previous studies on anxiety-like behavior in Fmr1KO mice revealed mixed results that are not always consistent with the enhanced emotionality observed in patients. Reduced anxiety-like behaviors were reported in the Fmr1KO using different behavioral tests such as the open field, the light-dark box or the elevated plus-maze (Liu and Smith, 2009; Jung et al., 2012; Busquets-Garcia et al., 2013). Other studies did not report this alteration in anxiety measurements (Mineur et al., 2002; Thomas et al., 2011), a discrepancy that might be due to specific experimental settings of the

study, including differences in strain, age, phase of the circadian cycle or lighting conditions (Spencer et al., 2011).

Finally, self-injurious behavior is another important trait of FXS (Tranfaglia, 2011), a feature that could be related to a reduction in pain sensitivity (Symons et al., 2010). Accordingly, the Fmr1KO shows a decreased nociceptive sensitization in models of inflammatory pain (Price et al., 2007; Busquets-Garcia et al., 2013). In addition, FMRP is expressed by nociceptors and localized in pain-sensing neurons, as well as in regions implicated in nociceptive control (Price et al., 2007). However, these findings have not been directly associated to the self-injurious behavior observed in humans, since Fmr1KO mice do not demonstrate such a behavior. Nevertheless, it is reasonable that an elevated pain threshold or alterations in pain pathways could underlie the persistence of self-injurious behavior (Peebles et al., 2012). Although more studies are warrant, evidence showed that self-injurious behavior may be modulated by increasing pain sensitivity in neurodevelopmental disability patients (Symons et al., 2004), and this could be one of the therapeutic goals in FXS.

## New insights into the cellular and molecular alterations

Fragile X Mental Retardation Protein acts as an inhibitor of local translation for specific mRNAs in neuronal dendritic spines (Darnell et al., 2011). The most frequent cellular phenotype associated with the loss of FMRP in rodents is an aberrant increase in the immature dendritic protrusions or filopodia (Hinton et al., 1991; Grossman et al., 2006). In agreement, alterations in neuronal spine density and morphology are *post-mortem* features of neurons in different brain areas of FXS patients (He and Portera-Cailliau, 2013). The altered dendritic spine number and morphology/functionality may underlie the hyperexcitability of Fmr1KO neuronal

circuits, which may play a relevant role in the pathophysiology of the disease (Gibson et al., 2008). Similarly, other neurological disorders, including epilepsy, tuberous sclerosis complex or Rett syndrome, show hyperexcitable neuronal circuits resulting from the unbalance between excitatory and inhibitory inputs (Eichler and Meier, 2008; Bateup et al., 2013). The structural abnormalities of synapses in Fmr1KO mice are associated to changes in functional synaptic connectivity (Pfeiffer and Huber, 2009), revealing the significance of FMRP in regulating neuronal and spine development, and the possibility of targeting this structural dysfunction pharmacologically (Dolan et al., 2013).

Synaptic transmission has been found altered in Fmr1KO mice. Long-term potentiation (LTP) and long-term depression (LTD) are phenomena widely accepted to underlie synaptic changes during learning and memory (Lynch, 2004; Massey and Bashir, 2007). Exaggerated hippocampal LTD, and specifically that dependent on metabotropic glutamate receptor (mGluR) activation (mGluR-LTD) has been reproducibly reported in Fmr1KO mice (Huber et al., 2002; Zhang et al., 2009). In this case, the lack of mRNA repression by FMRP in the Fmr1KO, increases the translation of different mRNAs necessary for LTD and enhances the internalization of AMPA receptors (Waung et al., 2008). The supposition that particular FXS symptoms could be explained by excessive protein synthesis downstream of over-active mGluR signaling led to the proposal entitled "mGluR theory of fragile X" (Bear and Huber, 2004). This was followed by the design of pharmacological approaches targeting mGluR5 (Krueger and Bear, 2011). Interestingly, the antagonists of mGluR5 CTEP was effective in the normalization of the Fmr1KO phenotype (Michalon et al., 2012; Michalon et al., 2014). Indeed, chronic CTEP administration normalized ERK and mTOR activity in Fmr1KO mice, and corrected the altered hippocampal GluR-LTD,

the seizure susceptibility, the deficits in memory and the in FXS, modifying to some extend the activity of brain regions such as the amygdala, the hypothalamus and the hippocampus (Michalon et al., 2012; Michalon et al., 2014). Interestingly, several mGlu5 inhibitors are under clinical examination in FXS: RO4917523 (Hoffmann-La Roche), STX107 (Seaside Therapeutics) and mavoglurant/AFQ056 (Novartis), although this last one has been recently called off (FRAXA, personal communication). It will be of great interest to analyze the results of these clinical trials to better design clinical tests for this and other therapeutic strategies (Gomez-Mancilla et al., 2014).

The studies in hippocampal LTP found either reduced (Hu et al, 2008; Lauterborn et al, 2007; Lee et al, 2011; Shang et al, 2009), or no major alterations in this parameter (Auerbach and Bear, 2010; Bear et al, 2004; Godfraind et al, 1996; Paradee et al, 1999), maybe due to the use of different induction protocols. The deficient LTP, specifically NMDA-dependent LTP, was related to a selective impairment of the signal transduction between the small GTP-ase Ras and the phosphoinositide 3-kinase (PI3K), that affects AMPA-type ionotropic glutamate receptor trafficking to the synapse (Hu et al., 2008; Stornetta and Zhu, 2011; Lim et al., 2014). In this regard, the modulation of the Ras/PI3K signaling, observed after combined treatment with activating compounds for serotonin 5HT2B and dopamine D1 receptors enhanced the Ras/PI3K and recovered AMPA receptor-dependent plasticity, correlating with the restoration of associative learning (Lim al., 2014). The fact that this approach used FDA-approved psychoactive drugs, and that these drugs are commonly used to treat a number of mental and psychiatric disorders (Roth et al. 2004; Beaulieu, 2012), may accelerate its testing in FXS patients.

Another well-accepted theory to understand the pathophysiology of FXS involves an

alteration of the inhibitory GABAergic signaling (D'Hulst and Kooy, 2007). Different alterations in brain regions of the Fmr1KO involving the GABAergic system include, a decreased GABA receptor signaling efficacy, a down-regulation of tonic GABA receptor-mediated inhibition and morphological defects of GABA releasing interneurons (**Figure 1**) (Levenga et al., 2010; Paluszkiewicz et al., 2011). Interestingly, a GABAB agonist, baclofen, was found effective in preventing the epileptic phenotype in Fmr1KO mice (Pacey et al., 2009), and arbaclofen, (R-baclofen) corrected synaptic abnormalities and reduced the elevated AMPA receptor internalization in Fmr1KO to control values (Henderson et al., 2012).

Overall, the glutamatergic and the GABAergic theories propose an exaggerated excitatory mGluR signaling and a decreased GABA signaling, suggesting an excitatory/inhibitory unbalance that could underlie most of FXS central traits, and motivate the most advanced therapies under clinical studies. However, in this review we want to highlight interesting alternative theories that have appeared recently in the fragile X syndrome field.

Mammalian target of rapamycin pathway as an intracellular therapeutic strategy

FXS has also been related to alterations in the intracellular signaling of the mammalian target of rapamycin (mTOR) pathway (Sharma et al., 2010). The deregulation of mTOR has also been associated with other neurological disorders characterized by presenting cognitive deficiencies (Troca-Marin et al., 2012). mTOR pathway components are present at synapses and influence synaptic plasticity through the control of local protein synthesis (Tang and Schuman, 2002). mTOR is activated in dendrites by stimulation of group I mGluRs and is required for mGluR-LTD at CA1 hippocampal synapses (Hou and Klann, 2004). Several groups have recently analyzed the protein kinase B (Akt)/mTOR pathway in Fmr1KO mice (Gross et al.,

2010; Sharma et al., 2010; Busquets-Garcia et al., 2013). Thus, increased activities of phosphoinositide 3-kinase (PI3K), Akt, and mTOR (Figure 1) have been detected in cortical synaptoneurosomes and hippocampal lysates from Fmr1KO mice (Sharma et al., 2010; Gross et al., 2010) and post-mortem tissues from FXS patients (Hoeffer et al., 2012). Additionally, PI3K inhibition rescues the excess translation and subsequent AMPA receptor endocytosis revealed in the Fmr1KO mice (Gross et al., 2010). In contrast, when Ras/PI3K signaling was tested during active learning, the maximal PI3K signaling was reduced in Fmr1KO mice (Hu et al., 2008; Lim et al., 2014). Indeed, as the signaling dynamic range of PI3K could be more important than the absolute signaling level for the capacity of synaptic plasticity and learning it can not be discarded that drugs stimulating the PI3K-Ras cascade could have beneficial effects in FXS (Lim et al., 2014). Although more studies are needed, the overall deregulation of PI3K/Ras signaling and aberrant mTOR-dependent protein translation seem to crucially contribute to the symptomatic manifestations in FXS.

Interestingly, genetic reduction of an mTOR pathway component, p70 ribosomal S6 kinase 1 (S6K1) (**Figure 1**), prevents the molecular, cellular and behavioral phenotypes in Fmr1KO mice (Bhattacharya et al., 2012). At the pharmacological level, the mTOR inhibitor temsirolimus (Guertin and Sabatini, 2009) also normalized in Fmr1KO mice the cognitive deficit in the object recognition test and the susceptibility to audiogenic seizures (Busquets-Garcia et al., 2013). In summary, the genetic reduction of mTOR pathway components, the direct pharmacological blockade of the mTOR pathway or the modulation of PI3K/Ras cascade can prevent a broad range of phenotypes and this could lead to find new therapeutic options for FXS.

The endocannabinoid system: a new possible therapeutic target for FXS

Several of the behavioral and biochemical responses that are modified in FXS are modulated by the endocannabinoid system (Kano et al., 2009; Fride, 2005), an important neuromodulatory system regulating synaptic plasticity (Castillo et al., 2012). Indeed, the activation of CB1 receptors, the main cannabinoid receptor in the brain, produces dose-dependent deficits in cognitive performance, analgesic effects, modifies anxiety-like behavior and affect neuronal network excitability (Fride, 2005; Kano et al., 2009). Moreover, some of the pharmacological effects of cannabinoid agonists are mediated by the activation of the Akt/mTOR signaling pathway in the brain (Puighermanal et al., 2012). CB1 receptors are abundantly expressed at presynaptic GABAergic contacts and, to a lesser extent, on glutamatergic terminals (Marsicano and Lutz, 1999), where they modulate neurotransmitter release. Cannabinoid receptors. upon activation by their endogenous ligands (endocannabinoids) that are produced "on demand" after depolarization, may reduce presynaptic neurotransmitter release through feedback inhibition (Wilson and Nicoll, 2002). Interestingly, postsynaptic activation of mGluR5 is a key physiological mechanism that promotes the synthesis of endocannabinoids in response to synaptic activity (Varma et al., 2001).

The responses mediated by endocannabinoids on GABAergic synapses in the hippocampus and dorsal striatum after mGluR5 activation are enhanced in Fmr1KO mice (Zhang and Alger, 2010; Maccarrone et al., 2010). However, other responses to endocannabinoid, such as the LTD in excitatory terminals at ventral striatum and prefrontal cortex are abolished in Fmr1KO mice (Jung et al., 2012). These findings point to the possibility that defective endocannabinoid modulation of synaptic function in different brain areas may contribute to the symptoms of FXS.

Two therapeutic strategies apparently opposed and targeting the endocannabinoid system have been tested in the Fmr1KO model. A first strategy consisted in the enhancement of the main endocannabinoid in the brain, 2-arachidonoylglycerol (2-AG) (Jung et al., 2012) using JZL184, a specific inhibitor of the 2-AG degrading enzyme monoacylglycerol lipase (Long et al., 2009). This treatment increased brain 2-AG content and normalized both the anomalous synaptic plasticity in the prefrontal cortex and ventral striatum, and the hyperlocomotion and reduced anxiety-like behavior of Fmr1KO mice (Jung et al., 2012). A second strategy, based in the blockade of CB1 receptors (either pharmacologically or genetically), normalized the cognitive deficit, the enhanced seizure susceptibility, the decreased pain sensitivity and the abnormal mTOR signaling and dendritic spine morphology in the hippocampus of Fmr1KO mice (Busquets-Garcia et al., 2013). Both studies demonstrate the involvement of the endocannabinoid system in the pathophysiology of FXS, but further investigation is required to clarify the most beneficial therapeutic venue.

# Other experimental approaches to treat FXS

Additional therapeutical approaches have been tested in FXS based on the molecular pathophysiology of the disease (Hagerman et al., 2012). Among those, the use of the mood stabilizer lithium was found to improve seizure susceptibility in Fmr1KO mice (Min et al., 2009), probably through its inhibitory action on glycogen synthase kinase 3 (GSK3), found constitutively overactive in the Fmr1KO (Min et al., 2009; Franklin et al., 2014). Although its beneficial effects reversing some FXS traits, lithium exerts side effects that could prevent its therapeutic utilization. Minocycline, a tetracycline antibiotic with neuroprotective effects in neurodegenerative conditions (Domercq and

Matute, 2004) has been found to enhance the maturation of hippocampal dendritic spines and to improve anxiety-like and exploratory behaviors in Fmr1KO mice (Bilousova et al., 2009). Memantine is a non-competitive N-methyl-D-aspartate (NMDA) receptor antagonist widely prescribed for treatment of Alzheimer's disease (Rogawski and Wenk, 2003) that inhibits the Ca<sup>2+</sup> influx. Experimental evidence showed that memantine could restore dendritic spines to normal values in the Fmr1KO neurons (Wei et al., 2012). Another promising compound is acamprosate, a mixed agonist/antagonist at NMDA receptors and activator of GABA-A receptors with possible inhibitory effects at group I mGluRs (Mann et al., 2008). In humans, acamprosate was well tolerated and improved social behavior, reducing the inattention/hyperactivity in FXS patients (Erickson et al., 2013).

### **3- Conclusion**

At the present time, the treatment of patients with FXS is only symptomatic. These symptomatic treatments include psychostimulants that ameliorate attention deficit and hyperactivity, and selective serotonin reuptake inhibitors that reduce aggression and anxiety-like responses. New strategies for therapeutic intervention have been recently proposed based on the basic research presented above. These new findings reinforce the potential therapeutic value of drugs that modulate mGluR5, GABA<sub>B</sub>, mTOR or CB1 receptor function. The ongoing clinical trials targeting mGluR5 and GABA<sub>B</sub> receptors, and the possibility to initiate novel trials with mTOR or CB1 receptor modulators will soon validate the translational power of the animal models of FXS to predict the most suitable treatments in humans. Importantly, all these promising approaches are able normalize cognitive deficits together with other phenotypes of

FXS in the preclinical models and, if confirmed in humans, they could improve enormously the quality of life of FXS patients compared to the actual symptomatic treatments.

# Figure legend

**Figure 1.** Schematic diagram showing the main molecular alterations described in FXS and the possible therapeutic site of action of the principal therapeutic approaches currently under study.

Right panel: The physiological situation in control conditions is represented. FMRP regulates the translation of several transcripts important for synaptic plasticity (Darnell et al., 2011) among those MMP-9 (Janusz et al., 2013), contributing to the normal behavioral output. The activity of the mTOR pathway is modulated by the balance between the excitatory and inhibitory drives in principal excitatory neurons. Presynaptic GABA<sub>B</sub> receptors and CB1 receptors modulate neurotransmitter release. CB1 receptors are more abundant in GABAergic terminals than in glutamatergic terminals.

Left panel: In the fragile X syndrome condition, the uncontrolled over-activity of mGluR5 (Bear et al., 2004) and the reduced GABAergic transmission (Levenga et al., 2010) may shift the excitatory/inhibitory input in principal neurons leading to the uncontrolled over-activation of the mTOR pathway and the phosphorylation of different upstream or downstream targets such as phosho-Akt or phosho-p70S6K, respectively (Sharma et al., 2010; Busquets-Garcia et al., 2013). The Ras/PI3K/Akt has been found impaired during active learning in Fmr1KO (Lim et al., 2014). Serotonin and dopamine receptor-acting compounds manage to re-establish the Ras/PI3K signaling recovering normal synaptic plasticity and memory performance (Lim et al., 2014). Under FXS conditions, the activation of protein translation machinery is favored, leading to aberrant enhanced protein synthesis and synaptic plasticity, enhanced internalization of AMPA receptors, changes in the structure of

spines to immature appearance, decreased phosphorylation of GSK3 to enhance its kinase activity, and enhanced activity of MMP-9. Treatments acting on mGluR5 or the GABA<sub>B</sub> receptor agonist, CTEP and arbaclofen, respectively (Michalon et al., 2012; Henderson et al., 2012) were effective in normalizing specific Fmr1KO phenotypes. The pharmacological reduction of actin polymerization by FRAX486, a p21-activated kinase inhibitor, was also valuable (Dolan et al., 2013). The genetic or pharmacological attenuation of CB1 receptor signaling, and the inhibition of mTOR activity also improved the cognitive performance (Busquets-Garcia et al., 2013). Lithium and other GSK3 inhibitors were found effective in reducing GSK3 activity (Min et al., 2009; Franklin et al., 2014), while minocycline may decrease MMP-9 activity in the FXS condition (Bilousova et al., 2009)

## Acknowledgements

AB-G was recipient of a predoctoral fellowship (Ministerio de Educación y Cultura) and supported by "Investments for the future" Programme IdEx Bordeaux (ANR-10-IDEX-03-02, French National Research Agency). Related research on the subject was supported by grants from FRAXA Research Foundation (AO), Jérôme Lejeune Foundation (AO), Ministerio de Ciencia e Innovación (#BFU2012-33500 to AO, #SAF2011-29864 to RM); Instituto de Salud Carlos III (RD06/0001/0001 to RM); PLAN E (Plan Español para el Estímulo de la Economía y el Empleo); Generalitat de Catalunya (SGR-2009-00731 to RM); ICREA (Institució Catalana de Recerca i Estudis Avançats) Academia to RM.

## **Bibliography**

- Auerbach BD, Bear MF. Loss of the fragile X mental retardation protein decouples metabotropic glutamate receptor dependent priming of long-term potentiation from protein synthesis. J Neurophysiol. 2010 Aug;104(2):1047-51.
- Bailey DB Jr, Raspa M, Olmsted M, Holiday DB. Co-occurring conditions associated with FMR1 gene variations: findings from a national parent survey. Am J Med Genet A. 2008 Aug 15;146A(16):2060-9.
- Baker et al., Fmr1 knockout mice: a model to study fragile X mental retardation. The Dutch-Belgian Fragile X Consortium. Cell. 1994 Jul 15;78(1):23-33.
- Baker KB, Wray SP, Ritter R, Mason S, Lanthorn TH, Savelieva KV. Male and female Fmr1 knockout mice on C57 albino background exhibit spatial learning and memory impairments. Genes Brain Behav. 2010 Aug;9(6):562-74.
- Bateup HS, Johnson CA, Denefrio CL, Saulnier JL, Kornacker K, Sabatini BL. Excitatory/inhibitory synaptic imbalance leads to hippocampal hyperexcitability in mouse models of tuberous sclerosis. Neuron. 2013 May 8;78(3):510-22.
- Bear MF, Huber KM, Warren ST. The mGluR theory of fragile X mental retardation. Trends Neurosci. 2004 Jul;27(7):370-7.
- Beaulieu JM. A role for Akt and glycogen synthase kinase-3 as integrators of dopamine and serotonin neurotransmission in mental health. J Psychiatry Neurosci. 2012 Jan;37(1):7-16.
- Berry-Kravis E, Raspa M, Loggin-Hester L, Bishop E, Holiday D, Bailey DB. Seizures in fragile X syndrome: characteristics and comorbid diagnoses. Am J Intellect Dev Disabil. 2010 Nov;115(6):461-72.
- Bhattacharya A, Kaphzan H, Alvarez-Dieppa AC, Murphy JP, Pierre P, Klann E. Genetic removal of p70 S6 kinase 1 corrects molecular, synaptic, and behavioral phenotypes in fragile X syndrome mice. Neuron. 2012 Oct 18;76(2):325-37.
- Bilousova TV, Dansie L, Ngo M, Aye J, Charles JR, Ethell DW, Ethell IM. Minocycline promotes dendritic spine maturation and improves behavioural performance in the fragile X mouse model. J Med Genet. 2009 Feb;46(2):94-102.
- Busquets-Garcia A, Gomis-González M, Guegan T, Agustín-Pavón C, Pastor A, Mato S, Pérez-Samartín A, Matute C, de la Torre R, Dierssen M, Maldonado R, Ozaita A. Targeting the endocannabinoid system in the treatment of fragile X syndrome. Nat Med. 2013 May;19(5):603-7.
- Castillo PE, Younts TJ, Chávez AE, Hashimotodani Y. Endocannabinoid signaling and synaptic function. Neuron. 2012 Oct 4;76(1):70-81.
- Chuang SC, Zhao W, Bauchwitz R, Yan Q, Bianchi R, Wong RK. Prolonged epileptiform discharges induced by altered group I metabotropic glutamate receptor-mediated synaptic responses in hippocampal slices of a fragile X mouse model. J Neurosci. 2005 Aug 31;25(35):8048-55.
- Chudley AE, Hagerman RJ. Fragile X syndrome. J Pediatr. 1987 Jun;110(6):821-31.
- Cornish K, Turk J, Hagerman R. The fragile X continuum: new advances and perspectives. J Intellect Disabil Res. 2008 Jun;52(Pt 6):469-82.
- D'Hooge R, Nagels G, Franck F, Bakker CE, Reyniers E, Storm K, Kooy RF, Oostra BA, Willems PJ, De Deyn PP. Mildly impaired water maze performance in male Fmr1 knockout mice. Neuroscience. 1997 Jan;76(2):367-76.
- D'Hulst C, Kooy RF. The GABAA receptor: a novel target for treatment of fragile X? Trends Neurosci. 2007 Aug;30(8):425-31.
- Darnell JC, Van Driesche SJ, Zhang C, Hung KY, Mele A, Fraser CE, Stone EF,

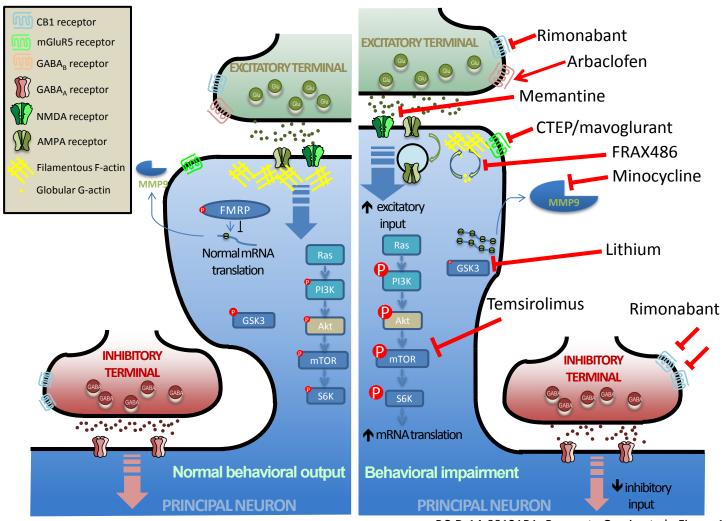
- Chen C, Fak JJ, Chi SW, Licatalosi DD, Richter JD, Darnell RB. FMRP stalls ribosomal translocation on mRNAs linked to synaptic function and autism. Cell. 2011 Jul 22;146(2):247-61.
- Dolan BM, Duron SG, Campbell DA, Vollrath B, Shankaranarayana Rao BS, Ko HY, Lin GG, Govindarajan A, Choi SY, Tonegawa S. Rescue of fragile X syndrome phenotypes in Fmr1 KO mice by the small-molecule PAK inhibitor FRAX486. Proc Natl Acad Sci U S A. 2013 Apr 2;110(14):5671-6.
- Dobkin, C., A. Rabe, R. Dumas, A. El Idrissi, H. Haubenstock and W. T. Brown (2000). "Fmr1 knockout mouse has a distinctive strain-specific learning impairment." Neuroscience 100(2): 423-429.
- Domercq M, Matute C. Neuroprotection by tetracyclines. Trends Pharmacol Sci. 2004 Dec;25(12):609-12.
- Eichler SA, Meier JC. E-I balance and human diseases from molecules to networking. Front Mol Neurosci. 2008 Mar 28;1:2.
- Erickson CA, Wink LK, Ray B, Early MC, Stiegelmeyer E, Mathieu-Frasier L, Patrick V, Lahiri DK, McDougle CJ. Impact of acamprosate on behavior and brain-derived neurotrophic factor: an open-label study in youth with fragile X syndrome. Psychopharmacology (Berl). 2013 Jul;228(1):75-84.
- Fisch GS, Simensen RJ, Schroer RJ. Longitudinal changes in cognitive and adaptive behavior scores in children and adolescents with the fragile X mutation or autism. J Autism Dev Disord. 2002 Apr;32(2):107-14.
- Franklin AV, King MK, Palomo V, Martinez A, McMahon LL, Jope RS. Glycogen synthase kinase-3 inhibitors reverse deficits in long-term potentiation and cognition in fragile X mice. Biol Psychiatry. 2014 Feb 1;75(3):198-206.
- Fride E. Endocannabinoids in the central nervous system: from neuronal networks to behavior. Curr Drug Targets CNS Neurol Disord. 2005 Dec;4(6):633-42.
- Gibson JR, Bartley AF, Hays SA, Huber KM. Imbalance of neocortical excitation and inhibition and altered UP states reflect network hyperexcitability in the mouse model of fragile X syndrome. J Neurophysiol. 2008 Nov;100(5):2615-26.
- Godfraind JM, Reyniers E, De Boulle K, D'Hooge R, De Deyn PP, Bakker CE, Oostra BA, Kooy RF, Willems PJ. Long-term potentiation in the hippocampus of fragile X knockout mice. Am J Med Genet. 1996 Aug 9;64(2):246-51.
- Gomez-Mancilla B, Berry-Kravis E, Hagerman R, von Raison F, Apostol G, Ufer M, Gasparini F, Jacquemont S. Development of mavoglurant and its potential for the treatment of fragile X syndrome. Expert Opin Investig Drugs. 2014 Jan;23(1):125-34.
- Gonçalves JT, Anstey JE, Golshani P, Portera-Cailliau C. Circuit level defects in the developing neocortex of Fragile X mice. Nat Neurosci. 2013 Jul;16(7):903-9.
- Gross C, Nakamoto M, Yao X, Chan CB, Yim SY, Ye K, Warren ST, Bassell GJ. Excess phosphoinositide 3-kinase subunit synthesis and activity as a novel therapeutic target in fragile X syndrome. J Neurosci. 2010 Aug 11;30(32):10624-38.
- Grossman AW, Elisseou NM, McKinney BC, Greenough WT. Hippocampal pyramidal cells in adult Fmr1 knockout mice exhibit an immature-appearing profile of dendritic spines. Brain Res. 2006 Apr 21;1084(1):158-64.
- Guertin DA, Sabatini DM. The pharmacology of mTOR inhibition. Sci Signal. 2009 Apr 21;2(67):pe24.
- Hagerman RJ. Lessons from fragile X regarding neurobiology, autism, and neurodegeneration. J Dev Behav Pediatr. 2006 Feb;27(1):63-74.

- Hagerman R, Lauterborn J, Au J, Berry-Kravis E. Fragile X syndrome and targeted treatment trials. Results Probl Cell Differ. 2012;54:297-335.
- Hayashi ML, Rao BS, Seo JS, Choi HS, Dolan BM, Choi SY, Chattarji S, Tonegawa S. Inhibition of p21-activated kinase rescues symptoms of fragile X syndrome in mice. Proc Natl Acad Sci U S A. 2007 Jul 3;104(27):11489-94.
- Hays SA, Huber KM, Gibson JR. Altered neocortical rhythmic activity states in Fmr1 KO mice are due to enhanced mGluR5 signaling and involve changes in excitatory circuitry. J Neurosci. 2011 Oct 5;31(40):14223-34.
- He CX, Portera-Cailliau C. The trouble with spines in fragile X syndrome: density, maturity and plasticity. Neuroscience. 2013 Oct 22;251:120-8.
- Henderson C, Wijetunge L, Kinoshita MN, Shumway M, Hammond RS, Postma FR, Brynczka C, Rush R, Thomas A, Paylor R, Warren ST, Vanderklish PW, Kind PC, Carpenter RL, Bear MF, Healy AM. Reversal of disease-related pathologies in the fragile X mouse model by selective activation of GABAB receptors with arbaclofen. Sci Transl Med. 2012 Sep 19;4(152):152ra128.
- Hinton VJ, Brown WT, Wisniewski K, Rudelli RD. Analysis of neocortex in three males with the fragile X syndrome. Am J Med Genet. 1991 Dec 1;41(3):289-94.
- Hoeffer CA, Sanchez E, Hagerman RJ, Mu Y, Nguyen DV, Wong H, Whelan AM, Zukin RS, Klann E, Tassone F.Altered mTOR signaling and enhanced CYFIP2 expression levels in subjects with fragile X syndrome. Genes Brain Behav. 2012 Apr;11(3):332-41.
- Hou L, Klann E. Activation of the phosphoinositide 3-kinase-Akt-mammalian target of rapamycin signaling pathway is required for metabotropic glutamate receptor-dependent long-term depression. J Neurosci. 2004 Jul 14;24(28):6352-61.
- Hu H, Qin Y, Bochorishvili G, Zhu Y, van Aelst L, Zhu JJ. Ras signaling mechanisms underlying impaired GluR1-dependent plasticity associated with fragile X syndrome. J Neurosci. 2008 Jul 30;28(31):7847-62.
- Huber KM, Gallagher SM, Warren ST, Bear MF. Altered synaptic plasticity in a mouse model of fragile X mental retardation. Proc Natl Acad Sci U S A. 2002 May 28;99(11):7746-50.
- Jacquemont S, Berry-Kravis E, Hagerman R, von Raison F, Gasparini F, Apostol G, Ufer M, Des Portes V, Gomez-Mancilla B. The challenges of clinical trials in fragile X syndrome. Psychopharmacology (Berl). 2013 Oct 31.
- Janusz A, Milek J, Perycz M, Pacini L, Bagni C, Kaczmarek L, Dziembowska M. The Fragile X mental retardation protein regulates matrix metalloproteinase 9 mRNA at synapses. J Neurosci. 2013 Nov 13;33(46):18234-41.
- Jung KM, Sepers M, Henstridge CM, Lassalle O, Neuhofer D, Martin H, Ginger M, Frick A, DiPatrizio NV, Mackie K, Katona I, Piomelli D, Manzoni OJ. Uncoupling of the endocannabinoid signalling complex in a mouse model of fragile X syndrome. Nat Commun. 2012;3:1080.
- Kano M, Ohno-Shosaku T, Hashimotodani Y, Uchigashima M, Watanabe M. Endocannabinoid-mediated control of synaptic transmission. Physiol Rev. 2009 Jan;89(1):309-80.
- Kooy RF. Of mice and the fragile X syndrome. Trends Genet. 2003 Mar;19(3):148-54.
- Krueger DD, Bear MF. Toward fulfilling the promise of molecular medicine in fragile X syndrome. Annu Rev Med. 2011;62:411-29.
- Lauterborn JC, Rex CS, Kramár E, Chen LY, Pandyarajan V, Lynch G, Gall CM. Brain-derived neurotrophic factor rescues synaptic plasticity in a mouse model of

- fragile X syndrome. J Neurosci. 2007 Oct 3;27(40):10685-94.
- Lee HY, Ge WP, Huang W, He Y, Wang GX, Rowson-Baldwin A, Smith SJ, Jan YN, Jan LY. Bidirectional regulation of dendritic voltage-gated potassium channels by the fragile X mental retardation protein. Neuron. 2011 Nov 17;72(4):630-42.
- Levenga J, de Vrij FM, Oostra BA, Willemsen R. Potential therapeutic interventions for fragile X syndrome. Trends Mol Med. 2010 Nov;16(11):516-27.
- Lim CS, Hoang ET, Viar KE, Stornetta RL, Scott MM, Zhu JJ. Pharmacological rescue of Ras signaling, GluA1-dependent synaptic plasticity, and learning deficits in a fragile X model. Genes Dev. 2014 Feb 1;28(3):273-89.
- Liu ZH, Smith CB. Dissociation of social and nonsocial anxiety in a mouse model of fragile X syndrome. Neurosci Lett. 2009 Apr 17;454(1):62-6.
- Long JZ, Li W, Booker L, Burston JJ, Kinsey SG, Schlosburg JE, Pavón FJ, Serrano AM, Selley DE, Parsons LH, Lichtman AH, Cravatt BF. Selective blockade of 2-arachidonoylglycerol hydrolysis produces cannabinoid behavioral effects. Nat Chem Biol. 2009 Jan;5(1):37-44.
- Lynch MA. Long-term potentiation and memory. Physiol Rev. 2004 Jan;84(1):87-136.
- Maccarrone M, Rossi S, Bari M, De Chiara V, Rapino C, Musella A, Bernardi G, Bagni C, Centonze D. Abnormal mGlu 5 receptor/endocannabinoid coupling in mice lacking FMRP and BC1 RNA. Neuropsychopharmacology. 2010 Jun;35(7):1500-9.
- Mann K, Kiefer F, Spanagel R, Littleton J. Acamprosate: recent findings and future research directions. Alcohol Clin Exp Res. 2008 Jul;32(7):1105-10.
- Markram K, Markram H. The intense world theory a unifying theory of the neurobiology of autism. Front Hum Neurosci. 2010 Dec 21;4:224.
- Marsicano G, Lutz B. Expression of the cannabinoid receptor CB1 in distinct neuronal subpopulations in the adult mouse forebrain. Eur J Neurosci. 1999 Dec;11(12):4213-25.
- Massey PV, Bashir ZI. Long-term depression: multiple forms and implications for brain function. Trends Neurosci. 2007 Apr;30(4):176-84.
- Michalon A, Sidorov M, Ballard TM, Ozmen L, Spooren W, Wettstein JG, Jaeschke G, Bear MF, Lindemann L. Chronic pharmacological mGlu5 inhibition corrects fragile X in adult mice. Neuron. 2012 Apr 12;74(1):49-56.
- Michalon A, Bruns A, Risterucci C, Honer M, Ballard TM, Ozmen L, Jaeschke G, Wettstein JG, von Kienlin M, Künnecke B, Lindemann L. Chronic metabotropic glutamate receptor 5 inhibition corrects local alterations of brain activity and improves cognitive performance in fragile X mice. Biol Psychiatry. 2014 Feb 1;75(3):189-97.
- Miller LJ, McIntosh DN, McGrath J, Shyu V, Lampe M, Taylor AK, Tassone F, Neitzel K, Stackhouse T, Hagerman RJ. Electrodermal responses to sensory stimuli in individuals with fragile X syndrome: a preliminary report. Am J Med Genet. 1999 Apr 2;83(4):268-79.
- Min WW, Yuskaitis CJ, Yan Q, Sikorski C, Chen S, Jope RS, Bauchwitz RP. Elevated glycogen synthase kinase-3 activity in Fragile X mice: key metabolic regulator with evidence for treatment potential. Neuropharmacology. 2009 Feb;56(2):463-72.
- Mineur YS, Sluyter F, de Wit S, Oostra BA, Crusio WE. Behavioral and neuroanatomical characterization of the Fmr1 knockout mouse. Hippocampus. 2002;12(1):39-46.

- Musumeci SA, Bosco P, Calabrese G, Bakker C, De Sarro GB, Elia M, Ferri R, Oostra BA. Audiogenic seizures susceptibility in transgenic mice with fragile X syndrome. Epilepsia. 2000 Jan;41(1):19-23.
- Pacey LK, Heximer SP, Hampson DR. Increased GABA(B) receptor-mediated signaling reduces the susceptibility of fragile X knockout mice to audiogenic seizures. Mol Pharmacol. 2009 Jul;76(1):18-24.
- Paluszkiewicz SM, Olmos-Serrano JL, Corbin JG, Huntsman MM. Impaired inhibitory control of cortical synchronization in fragile X syndrome. J Neurophysiol. 2011 Nov;106(5):2264-72.
- Paradee W, Melikian HE, Rasmussen DL, Kenneson A, Conn PJ, Warren ST. Fragile X mouse: strain effects of knockout phenotype and evidence suggesting deficient amygdala function. Neuroscience. 1999;94(1):185-92.
- Peebles KA, Price TJ. Self-injurious behaviour in intellectual disability syndromes: evidence for aberrant pain signalling as a contributing factor. J Intellect Disabil Res. 2012 May;56(5):441-52.
- Penagarikano O, Mulle JG, Warren ST. The pathophysiology of fragile x syndrome. Annu Rev Genomics Hum Genet. 2007;8:109-29.
- Penzes P, Cahill ME, Jones KA, VanLeeuwen JE, Woolfrey KM. Dendritic spine pathology in neuropsychiatric disorders. Nat Neurosci. 2011 Mar;14(3):285-93.
- Pfeiffer BE, Huber KM. The state of synapses in fragile X syndrome. Neuroscientist. 2009 Oct;15(5):549-67.
- Price TJ, Rashid MH, Millecamps M, Sanoja R, Entrena JM, Cervero F. Decreased nociceptive sensitization in mice lacking the fragile X mental retardation protein: role of mGluR1/5 and mTOR. J Neurosci. 2007 Dec 19;27(51):13958-67.
- Puighermanal E, Busquets-Garcia A, Maldonado R, Ozaita A. Cellular and intracellular mechanisms involved in the cognitive impairment of cannabinoids. Philos Trans R Soc Lond B Biol Sci. 2012 Dec 5;367(1607):3254-63.
- Rogawski MA, Wenk GL. The neuropharmacological basis for the use of memantine in the treatment of Alzheimer's disease. CNS Drug Rev. 2003 Fall;9(3):275-308.
- Roth BL, Sheffler DJ, Kroeze WK. Magic shotguns versus magic bullets: selectively non-selective drugs for mood disorders and schizophrenia. Nat Rev Drug Discov. 2004 Apr;3(4):353-9.
- Santoro MR, Bray SM, Warren ST. Molecular mechanisms of fragile X syndrome: a twenty-year perspective. Annu Rev Pathol. 2012;7:219-45.
- Shang Y, Wang H, Mercaldo V, Li X, Chen T, Zhuo M. Fragile X mental retardation protein is required for chemically-induced long-term potentiation of the hippocampus in adult mice. J Neurochem. 2009 Nov;111(3):635-46.
- Sharma A, Hoeffer CA, Takayasu Y, Miyawaki T, McBride SM, Klann E, Zukin RS. Dysregulation of mTOR signaling in fragile X syndrome. J Neurosci. 2010 Jan 13;30(2):694-702.
- Spencer CM, Alekseyenko O, Hamilton SM, Thomas AM, Serysheva E, Yuva-Paylor LA, Paylor R. Modifying behavioral phenotypes in Fmr1KO mice: genetic background differences reveal autistic-like responses. Autism Res. 2011 Feb;4(1):40-56.
- Spooren W, Lindemann L, Ghosh A, Santarelli L. Synapse dysfunction in autism: a molecular medicine approach to drug discovery in neurodevelopmental disorders. Trends Pharmacol Sci. 2012 Dec;33(12):669-84.
- Stornetta RL, Zhu JJ. Ras and Rap signaling in synaptic plasticity and mental disorders. Neuroscientist. 2011 Feb;17(1):54-78.

- Symons FJ, Thompson A, Rodriguez MC. Self-injurious behavior and the efficacy of naltrexone treatment: a quantitative synthesis. Ment Retard Dev Disabil Res Rev. 2004;10(3):193-200.
- Symons FJ, Byiers BJ, Raspa M, Bishop E, Bailey DB. Self-injurious behavior and fragile X syndrome: findings from the national fragile X survey. Am J Intellect Dev Disabil. 2010 Nov;115(6):473-81
- Tang SJ, Schuman EM. Protein synthesis in the dendrite. Philos Trans R Soc Lond B Biol Sci. 2002 Apr 29;357(1420):521-9.
- Thomas AM, Bui N, Graham D, Perkins JR, Yuva-Paylor LA, Paylor R. Genetic reduction of group 1 metabotropic glutamate receptors alters select behaviors in a mouse model for fragile X syndrome. Behav Brain Res. 2011 Oct 1;223(2):310-21
- Tranfaglia MR. The psychiatric presentation of fragile x: evolution of the diagnosis and treatment of the psychiatric comorbidities of fragile X syndrome. Dev Neurosci. 2011;33(5):337-48.
- Troca-Marín JA, Alves-Sampaio A, Montesinos ML. Deregulated mTOR-mediated translation in intellectual disability. Prog Neurobiol. 2012 Feb;96(2):268-82.
- Van Dam, D., R. D'Hooge, E. Hauben, E. Reyniers, I. Gantois, C. E. Bakker, B. A. Oostra, R. F. Kooy and P. P. De Deyn (2000). "Spatial learning, contextual fear conditioning and conditioned emotional response in Fmr1 knockout mice." Behav Brain Res 117(1-2): 127-136.
- Varma N, Carlson GC, Ledent C, Alger BE. Metabotropic glutamate receptors drive the endocannabinoid system in hippocampus. J Neurosci. 2001 Dec 15;21(24):RC188.
- Ventura R, Pascucci T, Catania MV, Musumeci SA, Puglisi-Allegra S. Object recognition impairment in Fmr1 knockout mice is reversed by amphetamine: involvement of dopamine in the medial prefrontal cortex. Behav Pharmacol. 2004 Sep;15(5-6):433-42.
- Wang T, Bray SM, Warren ST. New perspectives on the biology of fragile X syndrome. Curr Opin Genet Dev. 2012 Jun;22(3):256-63.
- Waung MW, Pfeiffer BE, Nosyreva ED, Ronesi JA, Huber KM. Rapid translation of Arc/Arg3.1 selectively mediates mGluR-dependent LTD through persistent increases in AMPAR endocytosis rate. Neuron. 2008 Jul 10;59(1):84-97.
- Wei H, Dobkin C, Sheikh AM, Malik M, Brown WT, Li X. The therapeutic effect of memantine through the stimulation of synapse formation and dendritic spine maturation in autism and fragile X syndrome. PLoS One. 2012;7(5):e36981.
- Wilson RI, Nicoll RA. Endocannabinoid signaling in the brain. Science. 2002 Apr 26;296(5568):678-82.
- Wondolowski J, Dickman D. Emerging links between homeostatic synaptic plasticity and neurological disease. Front Cell Neurosci. 2013 Nov 21;7:223.
- Zhang J, Hou L, Klann E, Nelson DL. Altered hippocampal synaptic plasticity in the FMR1 gene family knockout mouse models. J Neurophysiol. 2009 May;101(5):2572-80.
- Zhang L, Alger BE. Enhanced endocannabinoid signaling elevates neuronal excitability in fragile X syndrome. J Neurosci. 2010 Apr 21;30(16):5724-9.
- Zoghbi HY. Postnatal neurodevelopmental disorders: meeting at the synapse? Science. 2003 Oct 31;302(5646):826-30.



BC-D-14-00121R1- Busquets-Garcia et al., Figure 1